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Satoshi Ichikawa^a; Akira Matsuda^a

^a Graduate School of Pharmaceutical Sciences, Hokkaido University, Kita-ku, Sapporo, Japan

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Synthesis of Tunicaminyluracil Derivatives^{†,#}

Satoshi Ichikawa and Akira Matsuda*

Graduate School of Pharmaceutical Sciences, Hokkaido University, Kita-ku, Sapporo, Japan

ABSTRACT

A tunicaminyluracil derivative, which is a key component of the tunicamycin nucleoside antibiotics, was synthesized using a samarium diiodide (SmI₂) mediated aldol reaction and intramolecular Pummerer reaction as the key steps. The α -phenylthio ketone 11, the precursor of the samarium enolate, was prepared from D-galactose. Treatment of 11 with SmI₂ at -40° C resulted in complete conversion to the corresponding samarium enolate, and subsequent addition of uridine 5'-aldehyde 12 afforded the desired aldol products 13a,b. Compound 13a was converted to the sulfoxide 15 by a sequential diastereoselective reduction of the ketone and an oxidation with mCPBA. Activation of 15 with Tf₂O provided the desired cyclized compound 17. In this reaction, the aldol product 13a was also obtained as a consequence of a competitive intramolecular version of DMSO-oxidation via a 7-membered ring intermediate. Compound 18 or 19 are ready for use as a glycosyl donor

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in glycosylations to provide a range of analogues as potential glycosyltransferase inhibitors as well as related natural products.

Key Words: Tumicamycins; Tunicaminyluracil; Antibiotics; Samarium diiodide; Aldol reaction; Pummerer reaction.

INTRODUCTION

Tunicamycins^[1-6] (1, Figure 1), isolated from the fermentation broths of Streptomces lysosuperficus in 1971, are nucleoside antibiotics composed of uridine, N-acetylglucosamine (GlcNAc), an aminoundecose which is a unique higher carbon sugar called tunicamine, and an amide-linked fatty acyl side chain. [3-5] They exhibit a variety of biological properties including antibacterial, antiviral, antifungal, and antitumor activities.^[7-10] Treatment of eukaryotic cells with tunicamycins results in the complete truncation of the oligosaccharides from N-linked glycopeptides. Therefore, tunicamycins are also utilized as a biological tool to reveal functions of the oligosaccharides in the N-linked glycopeptides. Tunicamycins strongly and reversibly inhibits UDP-GlcNAc:polyprenol phosphate GlcNAc-1-P translocase, the enzyme which is responsible for the first N-acetylglucosamination of the N-linked glycopeptide in endothelial reticulum (ER). It is suggested that the structure of tunicamycins closely resemble the transition state of the transfer reaction of UDP-GlcNAc onto a dolicol monophosphate in the ER membrane catalyzed by the translocase (Figure 1). In particular, the C7'-C11' moiety of the tunicamycin structure forming a galactopyranoside can be considered as a mimic of the structure of a divalent metal chelated diphosphate. This structural mimicry has been utilized to design inhibitors of glycosyltransferases, which are responsible for oligosaccharide biosynthesis. [11,12] Thus, tunicaminyluracil (2), which lacks the GlcNAc moiety and the fatty acyl chain of the tunicamycins, would be expected to be a versatile synthetic intermediate for the synthesis of various glycosyltransferase inhibitors (for a recent review associated with glycosyltransferase inhibitors: Ref. [13]).

Their structural complexity also renders them worthy targets in organic synthesis. The total synthesis of tunicamycins has been accomplished by the groups of Suami's [14,15]

Figure 1. Structure of tunicamycins (1) and UDP-NacG1c.

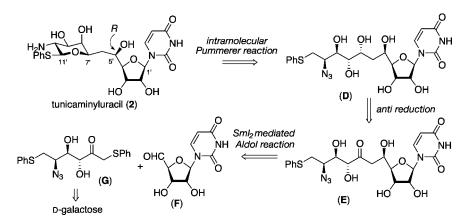
Tunicaminyluracil Derivatives

Scheme 1. Samarium diiodide (Sml₂)-mediated aldol reaction.

and Myers, $^{[16,17]}$ and other synthetic studies of related compounds have also been reported. $^{[18-23]}$

Previously, we applied the samarium diiodide (SmI₂) mediated C–C bond formation reaction to nucleoside chemistry^[24,25] and developed a novel aldol reaction via the samarium enolate ($\bf B$) generated by two electron reductions of the α -phenylthio ketone ($\bf A$) (Scheme 1).^[26] The neutral and mild reaction conditions and the reactivity of the regioselectively generated samarium enolate ($\bf B$) are suitable for the carbon-chain elongation to a base-labile nucleoside 5′-aldehyde derivative. This reaction was successfully applied to the total synthesis of the nucleoside antibiotic, herbicidin $\bf B$. This SmI₂-mediated aldol reaction could also be applied to the synthesis of tunicaminyluracil ($\bf 2$), the structure of which is a 5′-carbon-branched uridine derivative (Scheme 2). Here we describe the synthesis of tunicaminyluracil ($\bf 2$) as an extension of our SmI₂-mediated aldol reaction.

Our strategy includes the regioselective generation of a samarium enolate from the α -phenylthio ketone (**G**), aldol reaction with the uridine 5'-aldehyde (**F**) to assemble the undecose system (**E**). After stereoselective reduction of the 7'-keto group in **E**, cyclization of the resulting 7'-hydroxyl group in **D** to the carbon atom at the 11'-position by an intramolecular Pummerer reaction can be expected to give **2**. If such a cyclization occurs effectively, further introduction of certain carbohydrates using the resulting phenylthio pyranoside **2** as a versatile intermediate could be realized.



Scheme 2. Retro-synthetic analysis of tunicaminyluracil.

Therefore, this approach would provide a ready access to a range of sugar analogues for the development of glycosyltransferase inhibitors.

RESULTS AND DISCUSSION

The synthesis of the key α -phenylthio ketone 11 is summarized in Scheme 3. Protection of 2-azido-2-deoxy-3,4,6-tri-O-acetylgalactose 3^[28] with a TBDPS group gave only the β -galactoside 4, and successive removal of the acetyl groups followed by protection of the resulting secondary alcohols with an isopropylidene group under thermodynamic conditions provided 5. Introduction of a phenylthio group at the 6position, which would become a leaving group when the samarium enolate is generated, was conducted by activation of the hydroxyl group as its triflate, followed by displacement with thiophenol to afford 6 in 93% yield in 2 steps. After deprotection of the TBDPS group of 6 with tetrabutylammonium fluoride (TBAF), followed by reductive ring opening of the resulting pyranose by NaBH₄, the desired diol 7 was obtained in 86% yield. Subsequent protecting group manipulations afforded the monobenzoate 8 (83% yield for 3 steps) at the 5 position, and the remaining primary alcohol at position 1 was further converted to a phenylthio group in 90% yield as in the procedure for the preparation of 6. It should be noted that selective activation of the primary alcohol of 7 with Tf₂O followed by substitution with thiophenol was unsuccessful and gave a tetrahydropyran derivative as a result of ring closure of the triflate intermediate. Deprotection of the benzoyl group followed by Dess-Martin periodinane oxidation afforded the α-phenylthio ketone 11 without oxidizing either of the phenylthio groups in quantitative yield.

The key SmI₂-mediated aldol reaction was conducted (Scheme 4) and the results are summarized in Table 1. Our previous study revealed that the two-electron reduction

*Reagents and conditions; a. TBDPSCI, imidazole, DMF, 73%. b. i) NaOMe, MeOH. ii) 2,2'-dimethoxypropane, p-TsOH, acetone, 69% for 2 steps. c. i) Tf₂O, pyridine, CH₂Cl₂. ii) PhSH, Et₃N, CH₂Cl₂ 93% for 2 steps. d. i) TBAF, THF. ii) NaBH₄, MeOH, 86% for 2 steps. e. i) TBSCI, imidazole, DMF. ii) BzCI, pyridine. iii) TBAF, THF, 83% for 3 steps. f. i) Tf₂O, pyridine, CH₂Cl₂. ii) PhSH, Et₃N, CH₂Cl₂ 90% for 2 steps. g. NaOMe, MeOH, 90%.h. Dess-Martin periodinane, CH₂Cl₂, 99%.

Scheme 3. Preparation of α -phenylthio ketone.



OHC O NH 11, Sml₂ PhS N₃ O OH NH THF conditions O TBS TBSO OTBS 13a (5'R),13b (5'S)

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Scheme 4. Sml₂-mediated aldol reaction.

by SmI_2 to generate a samarium enolate from a 1-phenylthio-2-ulose derivative occurred at -78° C. However, the treatment of α -phenylthio ketone **11** with 2.2 equiv of SmI_2 followed by addition of 1.0 equiv of the aldehyde **12**^a at -78° C gave the desired aldol products **13a,b** in 13% yield (Table 1, entry 1, 5'R/5'S = 64/36). The low yield of the products and the large amount of the unreacted **11** observed in the reaction mixture indicated that the α -phenylthio ketone **11** without a hetero atom adjacent to the phenylthio group is less reactive to the two-electron reduction than that with an oxygen atom. After several attempts to optimize the reaction temperature, the reaction at -40° C gave complete consumption of **11**. Addition of the aldehyde **12** at -78° C after generation of the samarium enolate provided **13a,b** in 71% (entry 3, 5'R/5'S = 66/34), although the addition at 0° C resulted in inverted selectivity (entry 2, 5'R/5'S = 28/72). It is known that $5mI_2$ can reduce an azido group to the corresponding amino group. However, no such reduction of the azido group in **13** was detected, and therefore the two-electron reduction was chemoselectively accomplished.

Stereoselective reduction of the ketone 13a was required in the next step. Intramolecular hydride delivery from NaBH(OAc)₃ via a 6-membered transition state selectively afforded the desired 1,3-anti-diol 14 in quantitative yield. If the resulting hydroxyl group at the 7' position cyclized with the carbon atom at the 11 position to form a hexopyranose via an intramolecular Pummerer reaction, the desired tunicaminyluracil derivative 17 could be obtained. There are several methods available to promote the Pummerer reaction including the direct activation of a sulfide or of the corresponding sulfoxide. Direct activation of the sulfide 14 by treatment with NIS in the presence of a catalytic amount of TfOH resulted in the iodo-etherification product 16 between the 5'-hydroxyl group and the 5,6-double bond within the uracil base, and the desired cyclization failed to occur. Next, the activation of the corresponding sulfoxide 15 was examined (Ref. [29] a review for intramolecular Pummerer reaction: Ref. [30]). Oxidation of 14 with mCPBA provided the corresponding sulfoxide 15 as a mixture of diastereomers. An initial effort to activate 15 with (CF₃CO)₂O resulted in extensive trifluoroacetylation of the alcohols and only a trace amount of the desired product 17 was obtained. However, treatment of the sulfoxide 15 with Tf₂O in the presence of pyridine at -20° C provided the desired product 17 along with 13a as an inseparable mixture in a ratio of 62:38. After the mixture was treated with NaBH(OAc)₃, compound 17 could be separated from the corresponding reduced

^aThe uridine 5'-aldehyde derivative **12** was prepared by Dess-Martin periodinane oxidation of 2',3'-di-*O*-TBS uridine.

Table 1.

| | Temp. (°C)* | | | |
|-------|-------------|-----|-----------|-----------------|
| Entry | A | В | Yield (%) | Ratio (13a/13b) |
| 1 | -78 | -78 | 13 | 64/36 |
| 2 | -40 | 0 | 62 | 28/72 |
| 3 | -40 | -78 | 71 | 66/34 |

^{*}A: temperature at the addition of 11. B: temperature at the addition of 12.

product 14 by the usual silica gel column chromatography. The phenylthio glycoside 17 was a single β -anomer. The stereochemistry of 17 was determined by an NOE (3.6%) between H7' and H11' (Scheme 5).

Sulfonylation of the sulfoxide with Tf_2O promotes β -elimination to give a thiocarbenium intermediate. Intramolecular nucleophilic attack of the 7'-hydroxyl group on the 11'-carbon atom affords the cyclized product 17 (Scheme 6, path a). We suppose

*Reagents and conditions; a. NaBH₄, AcOH–CH₂Cl₂, 92%. b. *m*CPBA, CH₂Cl₂, 96%. c. with 14. NIS, TfOH, CH₂Cl₂. d. i) Tf₂O, pyridine, CH₂Cl₂. ii) NaBH₄, AcOH–CH₂Cl₂, 53% for 17, 36% for 14. e. Ac₂O, DMAP, CH₂Cl₂, 96%. f. i) PhSeH, Et₃N. ii) phthaloyl dichloride, DBU, toluene, 92% for 2 steps.

Scheme 5. Synthesis of tunicaminyluracil derivative.





Scheme 6. Reaction pathways to give 16 and 13a.

that the aldol product 13a is produced through the nucleophilic attack of the 7'hydroxyl group on the activated thionium cation with the formation of a 7-membered ring followed by hydrogen abstraction and elimination resulting in the oxidation of the alcohol, an intramolecular version of the DMSO-oxidation (Scheme 6, path b). The fact that the 5'-keto derivative or the sulfoxide of 13a, which is also possible a product if this reaction proceeds in an intermolecular fashion, was not detected in the reaction mixture indicates that the mechanism of the oxidation involves the intramolecular pathway b. Following acetylation of the 5'-hydroxyl group of 17, the 10'-azido group of the corresponding acetate 18 was reduced with PhSeH in the presence of Et₃N. The liberated amine was then protected with a phthaloyl group to afford 19, a fully protected tunicaminyluracil.

In conclusion, the tumicaminyluracil derivative 19 has been synthesized by an aldol reaction via the samarium enolate generated from the α-phenylthio ketone 11 and the intramolecular Pummerer reaction as the key steps. The SmI₂-mediated aldol reaction was successfully applied to the carbon-chain elongation of the uridine 5'-aldehyde derivative 12. Compound 18 or 19 would then be ready for use as glycosyl donor in glycosylations to provide a range of sugar analogues as well as related natural products.

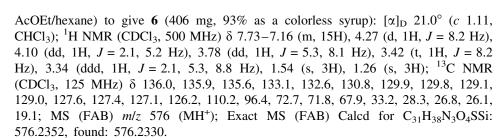
EXPERIMENTAL

General methods. Physical data were measured as follows: ¹H and ¹³C NMR spectra were recorded at 500 MHz and 125 MHz instruments in CDCl₃ as the solvent with tetramethylsilane as an internal standard. Chemical shifts are reported in parts per million (δ), and signals are expressed as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), or br (broad). All exchangeable protons were detected by addition of D₂O. Assignment of ¹H signals was based on two-dimentional NMR and NOE experiments. Mass spectra were measured on JEOL JMS-D300 spectrometer. TLC was done on Merck Kieselgel F254 precoated plates. Silica gel used for column chromatography was Merck silica gel 5715.

tert-Butyldiphenylsilyl 2-Azido-3,4,6-tri-*O*-acetyl-2-deoxy-β-D-galactopyranoside (4). A mixture of 3 (2.50 g, 7.50 mmol), TBDPSCl (2.34 mL, 9.00 mmol), and imidazole (1.23 g, 18.0 mmol) in DMF (40 mL) was stirred for 3 h at 50°C. After MeOH (5 mL) was added, the mixture was partitioned between AcOEt (200 mL) and H₂O (200 mL), and the organic layer was washed with H₂O (200 mL), brine (100 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 25% AcOEt/hexane) to give 4 (3.10 g, 73% as a colorless syrup): $[\alpha]_D - 9.29^\circ$ (*c* 0.96, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 7.73–7.36 (m, 10H), 5.23 (d, 1H, J = 3.0 Hz), 3.68 (dd, 1H, J = 3.3, 10.8 Hz), 4.46 (d, 1H, J = 7.7 Hz), 3.96 (dd, 1H, J = 6.6, 11.2 Hz), 3.89 (dd, 1H, J = 6.6, 11.2 Hz), 3.72 (dd, 1H, J = 7.6, 10.7 Hz), 3.53 (t, 1H, J = 6.6 Hz), 2.17 (s, 3H), 2.03 (s, 3H), 1.91 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 170.5, 170.3, 170.0, 136.1, 136.0, 133.1, 132.6, 130.3, 130.1, 127.9, 127.7, 97.2, 71.5, 70.8, 66.7, 63.7, 61.4, 27.0, 20.9, 20.8, 20.7, 19.4; MS (FAB) m/z 568 (MH⁺); Exact MS (FAB) Calcd for C₂₈H₂₉N₃O₈Si: 570.2271, found: 570.2291.

tert-Butyldiphenylsilyl 2-Azido-2-deoxy-3,4-O-isopropylidene-β-D-galactopyranoside (5). A mixture of 4 (2.84 g, 5.00 mmol) in MeOH (50 mL) containing NaOMe in MeOH (28%, 100 μL) was stirred for 2 h at room temperature. After neutralized by adding Dowex 50 (H⁺), the resin was removed by filtration, and the filtrate was evaporated under reduced pressure. The residue was coevaporated with toluene (3 × 10 mL). The residue and anhydrous p-TsOH (85 mg, 0.5 mmol) in acetone (50 mL) was stirred for 5 h at room temperature. The mixture was neutralized with saturated aqueous NaHCO₃, and the mixture was concentrated under reduced pressure. The residue was partitioned between AcOEt (200 mL) and H₂O (200 mL), and the organic layer was washed with H₂O (200 mL), brine (100 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by flush column chromatography (SiO₂, 25% AcOEt/hexane) to give 5 (3.10 g, 73% as a colorless syrup): [α]_D 52.8° (*c* 1.01, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 7.75–7.38 (m, 10H), 4.43 (d, 1H, J = 8.3 Hz), 3.91 (dd, 1H, J = 1.7, 5.2 Hz), 3.85 (dd, 1H, J = 5.4, 8.1 Hz), 3.66 (dd, 1H, J = 8.1, 11.9 Hz), 3.48 (dd, 1H, J = 5.6, 11.9 Hz), 3.43 $(t, 1H, J = 8.1 \text{ Hz}), 3.38 \text{ (ddd}, 1H, J = 1.7, 5.6, 8.1 \text{ Hz}), 1.56 \text{ (s, 3H)}, 1.30 \text{ (s, 3H)}; ^{13}\text{C}$ NMR (CDCl₃, 125 MHz) δ 137.4, 137.3, 135.2, 134.1, 131.7, 131.6, 129.4, 129.1, 112.3, 98.1, 79.2, 75.4, 74.5, 69.6, 63.7, 29.9, 28.4, 27.8, 20.6; MS (FAB) m/z 484 (MH⁺); Exact MS (FAB) Calcd for C₂₅H₃₄N₃O₅Si: 484.2267, found: 484.2251. Anal. Calcd for C₂₅H₃₃N₃O₅Si: C, 62.09; H, 6.88; N, 8.69. Found: C, 62.05; H, 6.89; N, 8.73.

tert-Butyldiphenylsilyl 2-Azido-2-deoxy-3,4-O-isopropylidene-6-S-phenyl-6-thio-β-D-galactopyranoside (6). A mixture of 5 (370 mg, 0.76 mmol) and Tf₂O (258 μL, 1.56 mmol) in CH₂Cl₂ (8 mL) containing pyridine (129 μL, 1.72 mmol) was stirred for 5 min at -20° C. The mixture was diluted with CH₂Cl₂ (20 mL), washed with H₂O (50 mL), saturated aqueous NaHCO₃ (30 mL), and brine (100 mL), dried (Na₂SO₄), and evaporated under reduced pressure. A solution of PhSH (117 μL, 1.17 mmol) and Et₃N (214 μL, 1.56 mmol) in CH₂Cl₂ (8 mL) was added to the above residue in CH₂Cl₂ (5 mL). The mixture was stirred for 1 h at room temperature, diluted with CH₂Cl₂ (20 mL), and washed with H₂O (50 mL). The organic layer was evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 4%



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(2S,3R,4S,5R)-2-Azido-3,4-(dimethylmethylenedioxy)-6-phenylthio-1,5-hexanediol (7). A mixture of 6 (3.60 g, 6.26 mmol) and TBAF (1 M, 6.89 mL, 6.89 mmol) in THF (60 mL) was stirred for 30 min at -20° C and evaporated under reduced pressure. Sodium borohydride (950 mg, 25.0 mmol) was added to the above residue in MeOH (60 mL) at -20° C, and the mixture was stirred for 30 min. The mixture was evaporated under reduced pressure, and then the residue was coevaporated with MeOH $(3 \times 10 \text{ mL})$. The residue was partitioned between AcOEt (200 mL) and H₂O (200 mL), and the organic layer was washed with H₂O (200 mL) and brine (100 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO2, 33% AcOEt/hexane) to give 7 (1.83 g, 86% as a colorless syrup): $[\alpha]_D - 29.8^{\circ}$ (c 0.99, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 7.38 (d, 2H, J = 7.7 Hz), 7.29 (t, 2H, J = 7.5 Hz), 7.21 (t, 1H, J = 7.4 Hz), 4.31 (dd, 1H, J = 1.8, 6.8 Hz), 4.27 (t, 1H, J = 6.5 Hz), 3.80 (m, 2H), 3.65 (m, 2H), 3.17 (dd, 1H, J = 6.5, 13.7 Hz), 3.06 (dd, 1H, J = 6.8, 13.7 Hz), 2.83 (d, 1H, J = 6.7 Hz, exchanged with D_2O), 2.27 (t, 1H, J = 5.7 Hz, exchanged with D_2O), 1.56 (s, 3H), 1.36 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 135.2, 129.8, 129.3, 129.2, 128.6, 126.7, 126.5, 126.1, 108.9, 68.0, 63.0, 61.7, 38.2, 26.5; MS (FAB) m/z 340 (MH⁺); Exact MS (FAB) Calcd for C₁₅H₂₂N₃O₄S: 340.1331, found: 340.1339.

(2S,3R,4S,5R)-2-Azido-5-benzoyloxy-3,4-(dimethylmethylenedioxy)-6-phenyl**thio-1-hexanol (8).** A mixture of **7** (500 mg, 1.47 mmol), TBSCl (243 mg, 1.67 mmol), and imidazole (220 mg, 3.23 mmol) in DMF (20 mL) was stirred for 1 h at 0°C. After MeOH (5 mL) was added, the mixture was partitioned between AcOEt (100 mL) and H₂O (100 mL), and the organic layer was washed with H₂O (100 mL) and brine (50 mL), dried (Na₂SO₄), and evaporated under reduced pressure. BzCl (256 μL, 2.21 mmol) was added to the residue in pyridine (20 mL), and the mixture was stirred for 12 h at room temperature. After MeOH (1 mL) was added, the mixture was evaporated under reduced pressure. The residue was partitioned between AcOEt (100 mL) and H₂O (100 mL), and the organic layer was washed with H₂O (100 mL) and brine (50 mL), dried (Na₂SO₄), and evaporated under reduced pressure. A mixture of the residue and TBAF (1 M, 1.67 mL, 1.67 mmol) in THF (20 mL) was stirred for 2 h at 0°C. The mixture was evaporated under reduced pressure, and the residue was purified by column chromatography (SiO₂, 10% AcOEt/hexane) to give 8 (534 mg, 83% as a colorless syrup): $[\alpha]_D - 52.7^{\circ}$ (c 1.04, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 8.12–7.18 (m, 10H), 5.22 (ddd, 1H, J = 1.6, 5.2, 8.8 Hz), 4.70 (dd, 1H, J = 1.6, 6.4 Hz), 4.39 (m, 1H), 3.53 (m, 2H), 3.47 (dd, 1H, J = 5.2, 13.7 Hz), 3.26 (dd, 1H, J = 8.8, 13.7 Hz), 1.96 (t, 1H, J = 5.2 Hz, exchanged with D₂O), 1.67 (s, 3H), 1.44 (s, 3H); 13 C NMR (CDCl₃, 125 MHz) δ 166.7, 135.2, 133.7, 130.1, 130.0, 129.6, 129.3, 128.8,

126.7, 109.5, 75.1, 72.1, 62.9, 61.6, 33.8, 26.8, 25.6; MS (FAB) m/z 444 (MH⁺); Exact MS (FAB) Calcd for $C_{22}H_{26}N_3O_5S$: 444.1593, found: 444.1580.

(2S,3R,4S,5R)-2-Azido-5-benzoyloxy-3,4-(dimethylmethylenedioxy)-1,6di(phenylthio)hexane (9). A mixture of 8 (570 mg, 1.28 mmol) and Tf₂O (261 μL, 1.53 mmol) in CH₂Cl₂ (15 mL) containing pyridine (129 μL, 1.72 mmol) was stirred for 5 min at -20°C, and diluted with CH₂Cl₂ (20 mL). The mixture was washed with H₂O (50 mL), saturated aqueous NaHCO₃ (30 mL), and brine (100 mL), dried (Na₂SO₄), and evaporated under reduced pressure. A mixture of the residue in CH₂Cl₂ (15 mL), PhSH (394 μ L, 3.84 mmol), and Et₃N (720 μ L, 5.12 mmol) was stirred for 1 h at room temperature, diluted with CH₂Cl₂ (20 mL), and washed with H₂O (50 mL). The organic layer was evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 4% AcOEt/hexane) to give 9 (614 mg, 90% as a colorless syrup): [α]_D 23.9° (c 1.02, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 8.03–7.12 (m, 15H), 5.07 (ddd, 1H, J = 3.0, 4.8, 8.3 Hz), 4.62 (dd, 1H, J = 3.0, 6.5 Hz), 4.41 (t, 1H, J = 6.8 Hz), 3.45 (dd, 1H, J = 6.8, 12.3 Hz), 3.41 (dd, 1H, J = 4.9, 13.8 Hz), 3.17 (dd, 1H, J = 8.4, 13.8 Hz), 2.97 (dd, 1H, J = 5.3, 13.8 Hz), 2.92 (dd, 1H, J = 7.0, 13.7 Hz), 1.61 (s, 3H), 1.39 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 166.2, 135.0, 134.5, 133.5, 131.2, 130.1, 129.9, 129.8, 129.5, 129.4, 128.7, 127.6, 126.9, 109.7, 78.3, 75.1, 71.7, 59.5, 36.9, 33.8, 25.7, 25.4; MS (FAB) m/z 536 (MH⁺); Exact MS (FAB) Calcd for C₂₈H₃₀N₃O₄S₂: 536.1677, found: 536.1670.

(2S,3R,4S,5R)-2-Azido-3,4-(dimethylmethylenedioxy)-1,6-di(phenylthio)hexane (10). A mixture of 9 (4.70 g, 8.74 mmol) in MeOH (90 mL) containing NaOMe in MeOH (28%, 0.8 mL) was stirred for 1 h at room temperature. After neutralized by adding Dowex 50 (H⁺), the resin was removed by filtration, and the filtrate was evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 4% AcOEt/hexane) to give 10 (3.40 g, 90% as a colorless syrup): $[\alpha]_D - 14.5^{\circ}$ (c 1.11, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 7.37–7.20 (m, 15H), 4.32 (dd, 1H, J = 4.8, 6.9 Hz), 4.26 (dd, 1H, J = 2.8, 6.9 Hz), 3.68 (ddd, 1H, J = 2.9, 6.4, 9.1 Hz), 3.57 (dd, 1H, J = 6.4, 11.5 Hz), 3.06 (m, 3H), 2.98 (dd, 1H, J = 6.5, 13.7 Hz), 1.51 (d, 1H, J = 5.8 Hz, exchanged with D₂O), 1.54 (s, 3H), 1.33 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 135.3, 134.7, 131.2, 130.1, 129.5, 129.4, 127.6, 127.0, 126.3, 109.3, 68.1, 60.6, 59.6, 38.5, 36.9, 26.6, 25.0, 21.3; MS (FAB) m/z 432 (MH⁺); Exact MS (FAB) Calcd for C₂₁H₂₆N₃O₃S₂: 432.1415, found: 432.1422.

(3R,4R,5S)-5-Azido-3,4-(dimethylmethylenedioxy)-1,6-di(phenylthio)-2-hexanone (11). A mixture of 10 (1.20 g, 2.79 mmol) and Dess-Martin periodinane (1.79 g, 4.19 mmol) in CH₂Cl₂ (30 mL) was stirred for 30 min at room temperature. After a mixture of saturated aqueous NaHCO₃ and saturated aqueous Na₂S₂O₃ (5:1, 50 mL) was added, the mixture was vigorously stirred until the organic layer turned to be clear. The organic layer was washed with H₂O (50 mL) and brine (50 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 8% hexane/AcOEt) to give 11 (1.20 g, 99%. as a colorless syrup): $[\alpha]_D$ 23.9° (c 1.17, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 7.40–7.19 (m, 15H), 4.69 (dd, 1H, J = 1.1, 8.7 Hz), 4.62 (d, 1H, J = 8.7 Hz), 4.19 (d, 1H, J = 15.7 Hz), 3.93 (d, 1H, J = 15.7 Hz), 3.34 (t, 1H, J = 6.6 Hz),3.27 (s, 2H), 1.60 (s, 3H), 1.32



(s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 204.4, 135.0, 134.4, 130.5, 130.2, 129.3, 129.0, 127.2, 126.9, 110.5, 79.7, 78.7, 57.7, 42.1, 34.8, 25.9, 23.8; MS (FAB) m/z 430 (MH⁺); Exact MS (FAB) Calcd for C₂₁H₂₄N₃O₃S₂: 430.1259, found: 430.1254.

1-[10-Azido-2,3-di-O-(tert-butyldimethylsilyl)-6,10,11-trideoxy-8,9-O-isopropylidene-11-S-phenyl-11-thio-L-lyxo-D-allo-undeculofuranosyl-(1,4)]uracil (13a) and 1-[10-Azido-2,3-di-O-(tert-butyldimethylsilyl)-6,10,11-trideoxy-8,9-O-isopropylidene-11-S-phenyl-11-thio-L-lyxo-L-talo-undeculofuranosyl-(1,4)]uracil (13b). Compound 11 (85 mg, 0.2 mmol) in THF (2 mL) was added dropwise to a THF solution of SmI_2 (0.1 M, 4.4 mL, 0.44 mmol) at $-40^{\circ}C$. After the TLC analysis indicated the disappearance of 11, oxygen gas was passed through the mixture. Then, 12 (94 mg, 0.2 mmol) in THF (2 mL) was added dropwise, and the mixture was stirred for 15 min at -78°C. After the mixture was allowed to warm to room temperature, saturated aqueous NH₄Cl was added. The mixture was filtrated through a Celite pad, and the filtrate was partitioned between AcOEt (50 mL) and H2O (50 mL), and the organic layer was washed with saturated aqueous NaHCO₃ (20 mL) and brine (20 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by flush column chromatography (SiO₂, 33% AcOEt/hexane) to give **13a** (74 mg, 47% as a white foam, fast moving) and 13b (38 mg, 24% as a white foam, slow moving).

For 13a: 1 H NMR (CDCl₃, 500 MHz) δ 8.95 (br s, 1H, exchanged with D₂O), 8.04 (d, 1H, J = 8.2 Hz), 7.43 (d, 2H, J = 7.7 Hz), 7.53 (t, 2H, J = 7.8 Hz), 7.26 (d, 1H, J = 7.7 Hz), 5.83 (d, 1H, J = 4.6 Hz), 5.69 (dd, 1H, J = 1.8, 8.1 Hz), 4.72 (d, 1H, J = 8.8Hz), 4.48 (d, 1H, J = 8.8 Hz), 4.31 (t, 1H, J = 4.5 Hz), 4.26 (d, 1H, J = 10.2 Hz), 3.93 (d, 1H, J = 4.2 Hz), 3.43 (d, 1H, J = 2.1 Hz), 3.33 (m, 3H), 3.29 (dd, 1H, J = 10.1, 19.3)Hz), 2.88 (dd, 1H, J = 1.9, 19.3 Hz), 1.62 (s, 3H), 1.31 (s, 3H), 0.92 (s, 9H), 0.90 (s, 9H), 0.11 (s, 6H), 0.07 (s, 6H); ¹³C NMR (CDCl₃, 125 MHz) δ 212.4, 163.6, 150.7, 141.4, 134.6, 130.7, 129.5, 127.5, 110.8, 102.3, 90.0, 87.1, 80.6, 79.2, 75.4, 72.4, 65.5, 57.9, 44.4, 35.2, 26.2, 26.1, 26.0, 24.0, 18.3, 18.2, -4.2, -4.4, -4.5, -4.6; MS (FAB) *m/z* 792 (MH⁺); Exact MS (FAB) Calcd for C₃₆H₅₈N₅O₉SSi₂: 792.3493, found: 792.3521.

For 13b: ¹H NMR (CDCl₃, 500 MHz) δ 8.87 (br s, 1H, exchanged with D₂O), 7.75 (d, 1H, J = 8.1 Hz), 7.41 (d, 2H, J = 7.6 Hz), 7.32 (t, 2H, J = 7.4 Hz), 7.26 (d, 1H, J = 7.4 Hz), 5.85 (d, 1H, J = 6.6 Hz), 5.73 (dd, 1H, J = 1.7, 8.1 Hz), 4.71 (d, 1H, J = 8.8 Hz, 4.47 (d, 1H, J = 8.8 Hz), 4.36 (dd, 1H, J = 4.5, 6.5 Hz), 4.29 (d, 1H, J = 11.1 Hz, 4.19 (dd, 1H, J = 1.4, 4.2 Hz), 3.86 (br s, 1H), 3.65 (d, 1H, J = 2.0 Hz), 3.29 (br s, 3H), 3.07 (d, 1H, J = 7.7 Hz), 2.76 (dd, 1H, J = 10.6, 18.7 Hz), 1.59 (s, 3H), 1.31 (s, 3H), 0.93 (s, 9H), 0.86 (s, 9H), 0.12 (s, 3H), 0.11 (s, 3H), 0.03 (s, 3H), -0.02 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 212.1, 163.4, 150.6, 141.8, 134.6, 130.8, 130.7, 129.5, 127.5, 110.8, 102.7, 89.6, 88.4, 80.5, 79.2, 74.8, 71.8, 67.8, 57.7, 43.7, 35.1, 26.1, 26.0, 24.0, 23.9, 18.3, 18.2, -4.1, -4.2, -4.4, -4.6; MS (FAB) m/z 792 (MH⁺); Exact MS (FAB) Calcd for C₃₆H₅₈N₅O₉SSi₂: 792.3493, found: 792.3486.

1-[10-Azido-2,3-di-O-(tert-butyldimethylsilyl)-6,10,11-trideoxy-8,9-O-isopropylidene-11-S-phenyl-11-thio-L-galacto-D-allo-undecofuranosyl-(1,4) luracil (14). Sodium borohydride (7.1mg, 189 μmol) was added to a mixture of AcOH and CH₂Cl₂ (1:2, 1 mL) at -20° C, and then a solution of **13a** (50 mg, 63 µmol) in CH₂Cl₂ (0.5 mL) was added dropwise to the mixture. After being stirred for 30 min at -20° C, the mixture was evaporated under reduced pressure, and the residue was coevaporated with

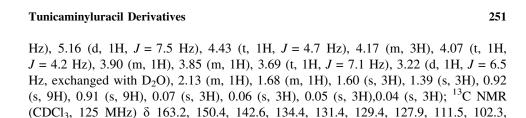
MeOH (3 \times 1 mL). The residue was partitioned between AcOEt (30 mL) and H₂O (20 mL), and the organic layer was washed with saturated aqueous NaHCO₃ (20 mL) and brine (100 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO2, 33% AcOEt/hexane) to give 14 (55 mg, quant. as a white foam): ^{1}H NMR (CDCl₃, 500 MHz) δ 9.49 (br s, 1H, exchanged with D_2O), 7.61 (d, 1H, J = 8.0 Hz), 7.42 (d, 2H, J = 7.8 Hz), 7.29 (t, 2H, J = 7.8 Hz, 7.21 (d, 1H, J = 7.7 Hz), 5.69 (d, 1H, J = 8.0 Hz), 5.69 (d, 1H, J = 5.7 Hz) Hz), 4.60 (t, 1H, J = 5.1 Hz), 4.46 (dd, 1H, J = 2.2, 6.4 Hz), 4.24 (ddd, 1H, J = 6.3, 9.6, 12.6 Hz), 4.13 (d, 2H), 4.08 (dd, 1H, J = 6.7, 9.4 Hz), 3.91 (d, 1H, J = 2.4 Hz), 3.68 (dd, 1H, J = 2.0, 7.1 Hz), 3.43 (d, 1H, J = 6.2 Hz), 3.33 (dd, 1H, J = 7.1, 13.6Hz), 3.27 (dd, 1H, J = 4.3, 13.6 Hz), 2.09 (ddd, 1H, J = 3.3, 3.8, 13.8 Hz), 1.70 (ddd, 1H, J = 1.8, 6.5, 13.8 Hz, 1.49 (s, 3H), 1.31 (s, 3H), 0.90 (s, 9H), 0.87 (s, 9H), 0.09 (s, 3H), 0.08 (s, 3H), 0.07 (s, 3H), 0.01 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 163.7, 150.8, 143.9, 135.2, 130.7, 129.3, 127.1, 109.3, 102.4, 94.8, 89.1, 78.7, 77.9, 73.2, 73.0, 67.7, 67.6, 58.6, 38.1, 36.5, 27.0, 26.1, 26.0, 25.1, 18.3, 18.2, -4.2, -4.4, -4.5, -4.7;MS (FAB) m/z 794 (MH⁺); Exact MS (FAB) Calcd for $C_{36}H_{60}N_5O_9SSi_2$: 794.3650, found: 794.3666.

1-[10-Azido-2,3-di-O-(tert-butyldimethylsilyl)-6,10,11-trideoxy-8,9-O-isopropylidene-11-phenylsulfinyl-L-galacto-D-allo-undecofuranosyl]uracil (15). A mixture of 14 (30 mg, 26 µmol) and mCPBA (4.6 mg, 26 µmol) in CH₂Cl₂ (5 mL) was stirred for 30 min at 0°C. After a mixture of saturated aqueous NaHCO₃ and saturated aqueous Na₂So₄0, (4:1, 5 mL) was added, the organic layer was washed with brine (5 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 66% AcOEt/hexane) to give a diastereomeric mixture of 15 (32 mg, quant. as a white foam): MS (FAB) m/z 810 (MH⁺); Exact MS (FAB) Calcd for C₃₆H₆₀N₅O₁₀SSi₂: 810.3599, found: 810.3598.

Thioglycoside 17. A mixture of **15** (7.0 mg, 8.7 μ mol) and Tf₂O (10.7 μ L, 52 μmol) in CH₂Cl₂ (100 μL) containing pyridine (6.6 μL, 104 μmol) was stirred for 5 min at -20° C, and diluted with CH₂Cl₂ (5 mL). The mixture was washed with H₂O (3 mL), saturated aqueous NaHCO3 (3 mL), and brine (3 mL), dried (Na2SO4), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 33% AcOEt/hexane) to give an inseparable mixture of 17 and 13a (5.7 mg, 89% as a glass, the ratio of 17 and 13a was 62:38 by ¹H NMR). To isolate pure 17, the next experiment was performed. NaBH₄ (9.4 mg, 189 µmol) was added to a mixture of AcOH and CH₂Cl₂ (1:2, 2 mL) at -20° C, and then the mixture of 17 and 13a (50 mg, 63 μmol, the ratio of 17 and 13a was 57:43) in CH₂Cl₂ (1 mL) was added dropwise to the above mixture. After being stirred for 30 min, the mixture was evaporated under reduced pressure, and the residue was coevaporated with MeOH $(3 \times 2 \text{ mL})$. The residue was partitioned between AcOEt (30 mL) and H₂O (20 mL), and the organic layer was washed with saturated aqueous NaHCO₃ (20 mL) and brine (10 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by flash column chromatography (SiO₂, 33% AcOEt/hexane) to give 17 (26 mg, 51.6% as a white foam, fast moving) and 14 (23 mg, 46% as a white foam, slow moving).

For 17: 1 H NMR (CDCl₃, 500 MHz) δ 8.68 (br s, 1H, exchanged with D₂O), 7.79 (d, 1H, J = 8.1 Hz), 7.51–7.30 (m, 5H), 5.70 (d, 1H, J = 8.1 Hz), 5.57 (d, 1H, J = 5.1





REPRINTS

C₃₆H₅₈N₅O₉SSi₂: 792.3493, found: 792.3496.

The physical data for **14** was in accordance with the compound obtained by reduction of **13a**.

92.4, 88.2, 86.7, 76.2, 74.3, 72.6, 69.2, 66.6, 64.0, 38.4, 27.8, 26.1, 26.0, 25.7, 18.3, 18.2, -4.2, -4.4, -4.5, -4.6; MS (FAB) *m/z* 792 (MH⁺); Exact MS (FAB) Calcd for

Acetate 18. A mixture of 17 (20 mg, 25 μmol), Ac₂O (2.8 μL, 30 μmol), Et₃N $(4.2 \mu L, 30 \mu mol)$, and DMAP (1 mg, 7.5 μ mol) in CH₃CN (1 mL) was stirred for 12 h at room temperature. After MeOH (1 mL) was added to the mixture, the mixture was evaporated under reduced pressure. The residue was partitioned between AcOEt (30 mL) and H₂O (20 mL), and the organic layer was washed with saturated aqueous NaHCO₃ (20 mL) and brine (10 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by flash column chromatography (SiO₂, 33%) AcOEt/hexane) to give 18 (21 mg, quant. as a white foam): ¹H NMR (CDCl₃, 500 MHz) δ 8.49 (br s, 1H, exchanged with D₂O), 7.78 (d, 1H, J = 8.1 Hz), 7.54–7.27 (m, 5H), 5.88 (d, 1H, J = 4.2 Hz), 5.74 (dd, 1H, J = 1.1, 8.1 Hz), 5.27 (ddd, 1H, J = 3.0, 5.8, 8.2 Hz), 5.07 (d, 1H, J = 7.4 Hz), 4.18 (dd, 1H, J = 3.2, 4.7 Hz), 4.15 (t, 1H, J = 5.1 Hz, 4.13 (t, 1H, J = 4.4 Hz), 4.04 (m, 1H), 4.03 (t, 1H, J = 4.3 Hz), 3.90 (t, 1H, J = 4.5 Hz), 3.67 (t, 1H, J = 6.9 Hz), 2.17 (ddd, 1H, J = 5.1, 8.3, 14.3 Hz), 2.10 (s 3H), 2.04 (ddd, 1H, J = 3.3, 8.2, 14.3 Hz), 1.55 (s, 3H), 1.37 (s, 3H), 0.88 (s, 9H), 0.87 (s, 9H), 0.09 (s, 3H), 0.06 (s, 3H), 0.05 (s, 3H),0.02 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 181.3, 169.6, 169.3, 163.0, 139.7, 134.6, 132.0, 129.3, 128.1, 111.4, 102.4, 101.7, 100.1, 88.6, 87.2, 84.9, 76.4, 76.2, 75.9, 71.9, 69.5, 68.4, 63.8, 36.0, 30.3, 27.9, 26.0, 21.4, 20.9, -4.0, -4.2, -4.5, -4.8; MS (FAB) m/z 883 (MH⁺); Exact MS (FAB) Calcd for $C_{38}H_{60}N_5O_{10}SSi_2$: 834.3599, found: 834.3578.

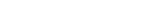
Protected tunicaminyluracil 19. A mixture of 18 (29 mg, 34 µmol) and PhSeH (10 μ L, 101 μ mol) in Et₃N (200 μ L) was heated for 1 h at 60°C. Additional PhSeH (21 μL, 202 μmol) was added to the mixture, and the mixture was further stirred for 1 h to complete the reaction. The mixture was evaporated under reduced pressure. A mixture of the residue, phthalic chloride (15 μL, 101 μmol), and DBU (32 μL, 217 μmol) in toluene (1 mL) was heated for 3 h at 100°C. The mixture was allowed to cool to room temperature and evaporated under reduced pressure. The residue was partitioned between AcOEt (20 mL) and H₂O (10 mL), and the organic layer was washed with brine (10 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, 33% AcOEt/hexane) to give 19 (31 mg, 98% as a pale yellow foam): ¹H NMR (CDCl₃, 500 MHz) δ 8.04 (br s, 1H, exchanged with D_2O), 7.87–7.17 (m, 10H), 5.92 (d, 1H, J = 4.8 Hz), 5.87 (d, 1H, J = 10.3 Hz), 5.76 (d, 1H, J = 8.2 Hz), 5.27 (ddd, 1H, J = 3.2, 5.4, 8.2 Hz), 4.95 (dd, 1H, J = 6.9, 10.3 Hz), 4.41 (t, 1H, J = 10.4 Hz), 4.33 (t, 1H, J = 7.3 Hz), 4.17 (t, 1H, J = 3.8 Hz), 4.14 (dd, 1H, J = 4.5, 8.1 Hz), 3.98 (t, 1H, J = 4.6 Hz), 3.93 (t, 1H, J = 4.1 Hz), 2.22 (ddd, 1H, J = 4.4, 8.0, 14.1 Hz), 2.15 (s, 3H), 2.11 (ddd, 1H, J = 5.5, 8.9, 14.1 Hz),

1.50 (s, 3H), 1.32 (s, 3H), 0.89 (s, 9H), 0.88 (s, 9H), 0.07 (s, 3H), 0.06 (s, 3H), 0.05 (s, 3H),0.03 (s, 3H); 13 C NMR (CDCl₃, 125 MHz) δ 169.5, 167.8, 162.6, 150.0, 139.5, 134.3, 131.7, 131.1, 128.9, 127.3, 132.6, 111.6, 102.4, 88.0, 85.2, 83.8, 75.7, 72.8, 72.0, 69.4, 68.8, 53.3, 36.0, 27.5, 25.8, 25.7, 21.2, 18.0, 17.9, -4.3, -4.5, -4.7, -4.9; MS (FAB) m/z 938 (MH⁺); Exact MS (FAB) Calcd for $C_{46}H_{64}N_3O_{12}SSi_2$: 938.3749, found: 938.3724.

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